

Serotonin receptors and their role in the pathophysiology and therapy of irritable bowel syndrome

C. Stasi · M. Bellini · G. Bassotti · C. Blandizzi · S. Milani

Received: 19 July 2013 / Accepted: 2 December 2013
© Springer-Verlag Italia 2013

Abstract

Background Irritable bowel syndrome (IBS) is a functional disorder of the gastrointestinal tract characterized by abdominal discomfort, pain and changes in bowel habits, often associated with psychological/psychiatric disorders. It has been suggested that the development of IBS may be related to the body's response to stress, which is one of the main factors that can modulate motility and visceral perception through the interaction between brain and gut (brain–gut axis). The present review will examine and discuss the role of serotonin (5-hydroxytryptamine, 5-HT) receptor subtypes in the pathophysiology and therapy of IBS.

Methods Search of the literature published in English using the PubMed database.

Results Several lines of evidence indicate that 5-HT and its receptor subtypes are likely to have a central role in the pathophysiology of IBS. 5-HT released from enterochromaffin cells regulates sensory, motor and secretory functions of the digestive system through the interaction with different receptor subtypes. It has been suggested that pain signals originate in intrinsic primary afferent neurons and are transmitted by extrinsic primary afferent neurons. Moreover, IBS is associated with abnormal activation of central stress circuits, which results in altered perception during visceral stimulation.

Conclusions Altered 5-HT signaling in the central nervous system and in the gut contributes to hypersensitivity in IBS. The therapeutic effects of 5-HT agonists/antagonists in IBS are likely to be due also to the ability to modulate visceral nociception in the central stress circuits. Further studies are needed in order to develop an optimal treatment.

C. Stasi (✉)

Department of Experimental and Clinical Medicine, University of Florence, Viale G.B. Morgagni, 85, 50134 Florence, Italy
e-mail: cristina.stasi@gmail.com

C. Stasi

Health Agency of Tuscany, Florence, Italy

M. Bellini

Gastroenterology Unit, Department of Gastroenterology, University of Pisa, Pisa, Italy

G. Bassotti

Gastroenterology and Hepatology Section, Department of Clinical and Experimental Medicine, University of Perugia, Perugia, Italy

C. Blandizzi

Department of Clinical and Experimental Medicine, University of Pisa, Pisa, Italy

S. Milani

Department of Biomedical, Experimental and Clinical Sciences, University of Florence, Florence, Italy

Keywords Irritable bowel syndrome · Serotonin · Serotonin receptors · Brain–gut axis

Introduction

Irritable bowel syndrome (IBS) is a multifactorial disorder, in which psychological abnormalities are significant factors contributing to both pathogenesis and clinical course. Serotonin (5-hydroxytryptamine, 5-HT) is likely to have a predominant role in the pathophysiology of IBS, through a variety of actions exerted in the central nervous system (CNS) and the enteric nervous system (ENS) [1, 2]. Dysfunctions of the central or peripheral serotonergic system can be involved in the pathophysiology of IBS, as suggested also by the therapeutic effects of both tricyclic

antidepressants and selective serotonin reuptake inhibitors [3].

Elevated plasma levels of 5-HT have been described in patients with diarrhea-predominant IBS (D-IBS), as compared with control subjects [4, 5]. Spiller et al. [6] reported an increased density of enterochromaffin cells (ECs) in rectal biopsies obtained from patients with D-IBS. In contrast, relatively low post-prandial plasma 5-HT levels have been detected in patients with constipation-predominant IBS (C-IBS) [7].

5-HT, released in the gut from ECs, regulates sensory, motor and secretory functions of the digestive system [8] through interactions with intrinsic and extrinsic nervous pathways. Intrinsic innervation to the gut is supplied by neurons of the ENS, including the myenteric and submucosal plexus. Extrinsic innervation is provided by the autonomic nervous system (both sympathetic and parasympathetic), and it is arranged to work in a bi-directional way: the brain can affect the ENS functions through the branches of the autonomous nervous system and, conversely, the gut can signal to the brain via extrinsic primary afferent neurons (EPANs), whose cell bodies are located in the ganglia of cranial nerves (e.g., ganglion nodosum and petrosum) or dorsal roots. Nociceptive signals are transmitted from abdominal viscera to specific laminae of the dorsal horns. Then, synaptic inputs activate specific second-order spinal neurons, leading to activation of specific brain areas. In parallel, ascending visceral nociceptive transmission is modulated by supraspinal structures (e.g., periaqueductal gray, raphe nucleus, locus coeruleus, thalamic regions), which either inhibit or facilitate nociceptive signaling through descending pathways.

Serotonergic, noradrenergic and dopaminergic fibers are the major components of these efferent descending projections. In this context, it has been suggested that visceral hypersensitivity in patients with IBS could be related to an abnormal signaling in the descending facilitatory pathway [9]. 5-HT has been found to exert opposite actions on intestinal motor activity and peristaltic reflex. On one hand, it can stimulate contractions through the release of acetylcholine from cholinergic neurons. On the other hand, it can facilitate enteric smooth muscle relaxation both by release of nitric oxide from nitrergic neurons and via a direct relaxation of smooth muscles [8].

The different gut effects of 5-HT are mediated by different serotonergic receptor subtypes located in the CNS [10], enteric neurons [11], gastrointestinal (GI) smooth muscle [12, 13] and secreting epithelial cells [14].

The present review article examines and discusses the role of 5-HT receptor subtypes in the pathophysiology and therapy of IBS.

5-HT receptors in the CNS and GI tract

5-HT exerts its biological activity through interaction with different receptors, currently classified into 7 groups on the basis of their structure, transduction mechanism and pharmacological profile [10, 14–16]: 5-HT_{1–7} (Table 1). Most of these receptors are expressed in the GI tract, and their stimulation plays different roles (either inhibitory or excitatory) in the control of intestinal motility and secretion. The 5-HT₃ receptor is coupled to an ion channel, whereas HT_{1,2,4,5,6,7} receptors are coupled to G proteins.

The class of 5-HT₁ receptors [17] is heterogeneous and includes several subtypes, such as 5-HT_{1A}, 5-HT_{1B}, 5-HT_{1C}, 5-HT_{1E}, 5-HT_{1F} and 5-HT_{1-like}. All 5-HT₁ receptor subtypes consist of a single peptide of variable length (from 374 to 421 amino acids), and they share at least 60 % homology in their transmembrane domains. 5-HT_{1A} receptors have a wide distribution in several brain regions involved in the modulation of emotions, such as the hippocampus, septum, dorsal raphe nuclei and amygdala [17], where they act mainly as inhibitory somatodendritic autoreceptors. However, at limbic level, particularly in the hippocampus, 5-HT_{1A} receptors are located post-synaptically, and here, their activation results in hyperpolarization of somatodendritic neuronal membrane [18]. These receptors have been found also in the neocortex and the gelatinous substance of the spinal cord, which are involved in the regulation of proprioceptive and integrative functions [17]. In the GI tract, 5-HT_{1A} receptors are expressed in the ENS, particularly in the submucosal and myenteric plexuses [11], where they mediate degranulation of enteric mast cells and release of mediators, including histamine [19].

5-HT_{1B} receptors are predominantly distributed in the striatum of basal ganglia and the prefrontal cortex, where they act as autoreceptors [18]. The 5-HT_{1C} subtype is similar in structure and transduction mechanism to receptors of the 5-HT₂ family, and for this reason, it has been renamed 5-HT_{2C} [20].

5-HT_{1D} receptors display a high degree of homology with 5-HT_{1B} receptors, but they are expressed with lower density. They inhibit neurotransmitter release [21] and mediate contraction of vascular smooth muscle cells [22]. The highest 5-HT_{1D} receptor densities are found in the raphe nuclei. 5-HT_{1D} receptors are expressed as α and β isoforms, endowed with similar pharmacological profiles [23–25]. The highest densities of 5-HT_{1E} receptor sites have been found in the caudate and putamen [26]. Their specific functional role is still not clear [27]. 5-HT_{1F} receptors have been identified in the CNS, particularly in the neocortex, where they might contribute to the integration of information associated with limbic functions [28]. 5-HT_{1-like} receptors are located in the CNS and intracranial

Table 1 Receptor subtypes in the brain and in the GI tract

Receptor subtypes	Location	Main functional role
5-HT _{1A}	CNS	Neuronal hyperpolarization [18]
	GI tract	Degranulation of enteric mast cells; release of mediators [19]
5-HT _{1B}	CNS	Autoreceptor; inhibition of neurotransmitter release [18]
5-HT _{1D}	CNS	Inhibition of neurotransmitter release [21]
	Intracranial vessels	Contraction of vascular smooth muscle [22]
5-HT _{1E}	CNS	Unknown [27]
5-HT _{1F}	CNS	Integration of sensorimotor or afferent information associated with limbic functions [28]
5-HT _{1-like}	CNS	Inhibition of noradrenaline release [29]
	Intracranial vessels	Smooth muscle contraction [30]
5-HT _{1P}	GI tract	Excitatory action on vagal afferent fibers [31]
5-HT _{2A}	CNS	Involvement into the neurochemical and behavioral effects of psychostimulants [33]
	GI tract	Contraction of gut smooth muscle [12]
5-HT _{2B}	GI tract	Increased response of the colonic longitudinal smooth muscle [38]
5-HT _{2C}	CNS	Production of regulation of emotional states [39]
	Choroid plexus	Cerebrospinal fluid [39]
5-HT ₃	CNS	Modulation of the release of other neurotransmitters such as dopamine, GABA, substance P and acetylcholine [42]
	GI tract	Motility [46] and pain transmission [8, 38]
5-HT ₄	CNS	Memory [47], cognitive function [48], affective symptoms [50]
	GI tract	Contraction of smooth colonic muscle
		Prokinetic effect Neurotransmitter release [31]
5-HT _{5A}	CNS	Regulation of affective states, learning, sensory perception, neuroendocrine functions and memory [54, 55]
5-HT ₆	CNS	Regulation of affective states [58]
5-HT ₇	CNS	Regulation of affective states
	GI tract	Muscle relaxant action of GI tract [59]

GI tract gastrointestinal tract, *5-HT* 5-hydroxytryptamine, *CNS* central nervous system, *GABA* gamma-aminobutyric acid

vessels. They inhibit noradrenaline release from sympathetic nerves [29] and vascular smooth muscle cell contraction [30]. 5-HT_{1P} receptors are expressed in the GI tract, where they mediate excitatory actions on vagal afferent fibers [31].

The 5-HT₂ receptor family includes three subtypes, named 5-HT_{2A}, 5-HT_{2B} and 5-HT_{2C}. They are expressed predominantly in peripheral tissues, such as stomach, intestine, heart and kidney [32], while in the CNS they are found in the cerebellum, lateral septum, hypothalamus and middle part of the amygdala. 5-HT_{2A} and 5-HT_{2C} receptors are known to mediate the neurochemical and behavioral effects of psychostimulants [33]. In the GI tract, they promote contractions of smooth muscle cells [12].

The 5-HT_{2B} receptor was first identified in the fundus of rat stomach [32]. Originally classified as 5-HT_{1-like} receptor [34], it has been recently assigned to the 5-HT₂ receptor family with the name of 5-HT_{2B} [20, 35, 36]. In the rat, 5-HT_{2B} receptor stimulation mediates hyperphagia and

reduces “grooming” behavior [37]. Peripherally, 5-HT_{2B} receptors are located in myenteric nerves and colonic smooth muscle cells, where they mediate contractile responses of the longitudinal muscle layer to electrical stimulation [38]. 5-HT_{2C} receptors are predominantly expressed in epithelial cells of the choroid plexus, cerebral cortex, hippocampus, amygdala, some components of basal ganglia, substantia nigra, substantia innominata and ventromedial hypothalamus [39].

5-HT₃ receptors belong to the ion-channel-linked receptor super-family, which includes nicotinic, cholinergic and gamma-aminobutyric acid (GABA) A receptors [40]. They are located in the hippocampus, dorsal motor nucleus of the solitary tract and area postrema [41]. At the CNS level, they are involved in the regulation of emetic responses to various stimuli, including anticancer chemotherapy. The activation of 5-HT₃ receptors elicits central effects comparable to those observed after administration of antipsychotic and anxiolytic drugs, due to their ability to modulate the release of other

neurotransmitters such as dopamine, GABA, substance P and acetylcholine [42]. In the GI tract, 5-HT₃ receptors are expressed in enteric neurons, smooth muscle cells, vagal and spinal primary afferent neurons, and in the spinal cord [43, 44]. 5-HT₃ receptors activate extrinsic sensory neurons which mediate pain [8, 38]. They also regulate the pacemaker activity of the interstitial cells of Cajal [45, 46].

5-HT₄ receptors are localized in the CNS, where it has been suggested they play a role in enhancing memory [47]. Several studies indicate that 5-HT₄ receptors are involved in cognitive functions [48], affective symptoms and the development of Alzheimer's disease [49, 50]. A study of Madsen et al. [51] showed that healthy women had a lower 5-HT₄ receptor binding in the limbic system. In the GI tract, 5-HT₄ receptors are expressed in enteric neurons and smooth muscle cells. After 5-HT₄ receptor activation, acetylcholine is released from enteric interneurons and motor neurons, thus promoting and maintaining propulsive motility [52]. In the heart, 5-HT₄ agonists elicit positive chronotropic and inotropic effects in the isolated human atrium [53].

5-HT_{5A} receptors are distributed predominantly in the cortex, hippocampus, hypothalamus, amygdala and cerebellum. Depending on their localization, 5-HT_{5A} receptors are involved in the regulation of several functions, such as the control of affective states, sensory perception and neuroendocrine functions [54]. The limbic distribution of these receptors suggests a role in learning, memory and emotional state [55, 56]. 5-HT_{5B} receptors are expressed in mice but not in humans [57].

5-HT₆ receptors are located in the striatum, amygdala, nucleus accumbens, olfactory tubercle and cortex. As demonstrated by several pharmacological studies, many antipsychotic drugs (clozapine, olanzapine and quetiapine) and antidepressants (clomipramine, amitriptyline and nortriptyline) act as high affinity antagonists of 5-HT₆ receptors [58].

5-HT₇ receptors are distributed in the limbic system and thalamocortical regions, where they are involved in the modulation of affective states. They are also expressed in smooth muscle cells of peripheral vessels and intestine, where they mediate muscle relaxation [59]. In this respect, Tonini et al. [60] found that 5-HT₇ receptors mediate intestinal smooth muscle relaxation and accommodation in the guinea pig ileum. The authors suggested that an abnormal stimulation of 5-HT₇ receptors may contribute to clinical syndromes, like IBS, and that they could be a possible candidate for therapeutic interventions.

Serotonergic transmission in gut–brain axis

5-Hydroxytryptamine is a very active mediator in both the ENS and the CNS. In the periphery, about 90 % of 5-HT is

synthesized in the gut by the ECs. Physiological intraluminal distension of the intestine and its propulsive activity promotes the release of 5-HT from ECs of the enteric mucosa [61]. Once released, 5-HT stimulates 5-HT₃ and 5-HT₄ receptors, located on intrinsic primary afferent neurons (IPANS) of the ENS, which mediate both secretory and motor responses. In the GI tract, both the secretory and peristaltic reflexes are regulated by 5-HT release from ECs and depend on the stimulation of 5-HT_{1B/1P} and 5-HT₄ receptors located on submucosal IPANS [8, 62]. Post-synaptic 5-HT₃ receptors are present in both enteric plexuses, particularly in the motor neurons that innervate smooth muscle [38].

Danzenbrink and Gebhart [63] demonstrated that 5-HT₁, 5-HT₂ and 5-HT₃ receptors mediate noxious visceral stimulus. A main function of 5-HT₃ receptors is the activation of EPANs, which mediate the pain and swelling of bowel wall associated with IBS [8, 38].

It has been suggested that pain signals originate in IPANS and are transmitted by EPANs [64]. The connections to the brain occur through vagal and spinal afferent nerves. Vagal afferent neurons have their cell bodies located in the nodose ganglia and project to the nucleus of the solitary tract, which in turn projects to the thalamus and transmits signals to the limbic system and to the frontal cortex for polymodal association and perception of pain.

First-order spinal afferent nerves synapse in the dorsal horn-lamina II and IV. Second-order neurones project to the brain through the spinothalamic and spinoreticular tract [65]. The second-order neurons synapse with third-order neurons in the thalamus and reticular nuclei. Spinal afferents transmit pain signals to the somatosensory cortex to discriminate and process pain information (Fig. 1).

Serotonergic fibers, descending from brain to the dorsal horn neurons, are responsible for visceral sensitivity, thus modulating perception during gut stimulation [66, 67].

Recently, Keszthelyi et al. [68] showed significantly lower mucosal and higher systemic concentrations of both 5-HT and kynurenic acid, a main kynurenine metabolite, in IBS patients as compared to healthy controls. Also, significant correlation between mucosal but not plasma concentrations of kynurenic acid and 5-HT and psychological state in IBS was observed [67]. Moreover, Stasi et al. [69], in diarrhea-predominant IBS (D-IBS), have shown a correlation between plasma cortisol and 5-HT, which could be explained by the concomitant activation of the hypothalamic-pituitary-adrenal (HPA) axis to limit local inflammatory processes in response to both exteroceptive and interoceptive stress (with the consequent activation of ECs, production of 5-HT, activation of mast cells and secretion of cytokines). These data suggest that an alteration of 5-HT production and consequently neurotransmission could be involved in psychological state and IBS symptoms.

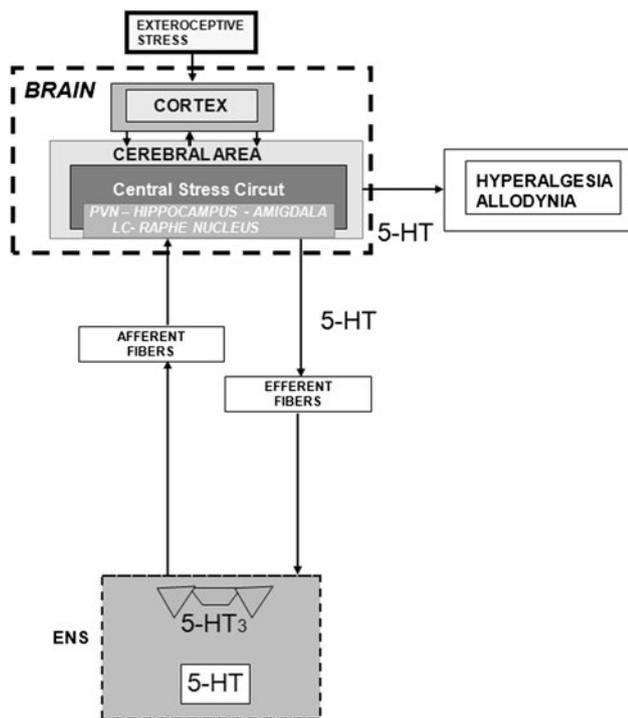


Fig. 1 (Modified from Stasi et al. [2]). Schematic model of serotonin-mediated brain–gut connection. In the ENS, 5-HT binds to 5-HT₃ receptors and via afferent fibers connects the gut to the central stress circuit (PVN, hippocampus, amygdala, LC, raphe nucleus and HPA axis). These cerebral areas are altered in IBS patients, because of exteroceptive chronic stress, leading to a local release of 5-HT. This may result in hyperalgesia and allodynia typical of this condition. The descending pathways may either inhibit or facilitate nociceptive signalling. 5-HT serotonin, ENS enteric nervous system, PVN paraventricular nucleus, LC locus coeruleus, HPA hypothalamic-pituitary-adrenal, IBS irritable bowel syndrome

IBS has a strong predominance in women. Reported prevalence rates of IBS range from 8 to 20 % in the general US population, with a 2:1 female-to-male ratio [70]. Nakai et al. [71] showed that 5-HT synthesis in the right medial temporal gyrus (multimodal sensory association cortex) was higher in female IBS patients than in female controls. This suggests a relationship with the pathological visceral pain processing of IBS female patients. It has been demonstrated that the number of 5-HT₄ receptors in the female brain is lower than in the male brain, suggesting that this might be the basis for the sex-specific difference in emotional control and the higher prevalence of affective diseases and visceral hypersensitivity in women [51].

Several factors, such as the high prevalence of anxiety and psychological disorders, have been shown to increase intestinal response to psychological stress [2]. Clinical response to serotonin drugs acting at the central level [72, 73] suggests an involvement of the limbic system in the pathophysiology of IBS. In recent years, the importance of the body's response to stress has repeatedly been

underlined and represents one of the main factors that can modulate motility and visceral perception through brain–gut interaction (brain–gut axis) [2, 9].

Irritable bowel syndrome is associated with visceral hypersensitivity, which can be related to deranged processing, representation and modulation of gut signals in the brain [74, 75]. Whitehead et al. [76] showed that patients with IBS have a significantly lower pain threshold than controls. Increased sensitivity to distention of the colon and rectum was observed in patients with D-IBS as compared with healthy subjects [77, 78], while studies in patients with C-IBS have yielded conflicting results [79, 80]. However, no significant differences in pain threshold have been shown in patients with D-IBS when compared with patients with C-IBS [81]. These data suggest that the alterations of 5-HT transmission in the CNS are similar in D- and C-IBS. Drossman et al. [82], using functional magnetic resonance imaging of the brain in a IBS patient, found a correlation between the severity of clinical symptoms and psychosocial state, with activation of the cingulate cortex, the critical center for pain control. Other studies have shown deactivation of the insula, the amygdala and the striatum of patients with IBS compared to controls [83, 84]. These brain areas are part of the central stress circuit, that is, under feedback control through the projections from the brainstem nuclei, in particular from serotonergic nuclei such as the raphe nucleus [2]. 5-HT receptors are expressed in cortical and limbic areas of the brain involved in emotional conditions and perception of visceral pain. Activation of 5-HT₃ receptors on the central terminals of spinal afferents increases the spinal transmission in the entire dorsal horn, which results in increased pain and reflex responses [85].

Moreover, stress may increase the permeability of the blood–brain barrier and enhance the action of some drugs on central targets when they are orally administered [86]. In animal models, transient blockade of the 5-HT₃ receptor by intrathecal injections of the antagonist ondansetron reduces mechanical allodynia after spinal cord injury [87]. Tegaserod antagonizes 5-HT_{2B} receptors at concentrations similar to those that activate 5-HT₄ receptors and has significant binding affinity for human recombinant 5-HT_{2A}, 5-HT_{2B} and 5-HT_{2C} receptors [88, 89]. The ability of tegaserod to alleviate abdominal pain and discomfort in patients with IBS is likely due not to its effects on peripheral 5-HT₄ receptors, but probably to its actions on 5-HT_{2A}, 5-HT_{2B} and 5-HT_{2C} receptors located in CNS.

Therapeutic approach to irritable bowel syndrome

Several studies have investigated alterations of serotonergic signaling in IBS; consequently, treatment strategies for

IBS involving 5-HT receptors are currently directed at managing the predominant symptoms. Initial pharmacological attempts have been primarily directed towards those receptors known to regulate intestinal function, such as 5-HT₁, 5-HT₃, 5-HT₄ and 5-HT₇ subtypes.

Ondansetron and alosetron are antagonists of 5-HT₃ receptors. They delay transit throughout the colon both in D-IBS and controls [90]. Moreover, ondansetron increases the consistency of feces both in healthy volunteers and IBS patients [91, 92].

Likewise, alosetron, another 5-HT₃ antagonist, was shown to be able to delay colonic transit in both healthy volunteers and D-IBS patients [5, 93].

Some studies suggest that antagonists of 5-HT₄ receptors may induce a significant slowing of intestinal transit, thus allowing a potential therapeutic benefit in patients with D-IBS [94]. Piboserod, an antagonist of 5-HT₄ receptors, showed no significant effects in the treatment of D-IBS [95]. In contrast, 5-HT₄ receptor agonists, such as tegaserod and prucalopride, accelerate GI transit in humans [96, 97].

In summary, these studies suggest that the antagonists of 5-HT₃ receptors can delay intestinal transit, while the agonists of 5-HT₄ receptors can lead to a transit acceleration.

Some patients with IBS display increased visceral sensitivity and/or an increased perception of intestinal distension. It has been observed that antagonists acting on 5-HT₃ receptors, expressed on post-synaptic neurons of the peripheral nervous system, can modify the visceral sensation [98], thus improving abdominal pain and discomfort.

Consistently with these findings, a recent multicenter, randomized study [99] has shown that ramosetron (5-HT₃ antagonist) can alleviate abdominal pain/discomfort and abnormal bowel habits in male patients with D-IBS. Previous experimental evidence showed that tegaserod (a partial 5-HT₄ receptor agonist) can induce a dose-dependent reduction in the discharge rate from mechanosensory neurons in the spinal cord of the cat [100]. A systematic review by Evans et al. [101] showed that in patients with C-IBS, tegaserod counteracted symptoms when compared with placebo, while it did not increase bowel movements. Since tegaserod was withdrawn from the market owing to cardiovascular adverse events, pharmacological research has been focused on the development of high-selective 5-HT₄ receptor agonists in an attempt to circumvent aspecific adverse effects.

Naronapride (ATI-7505) is a novel selective 5-HT₄ receptor agonist. In a clinical trial, the effects of this drug were evaluated after 9 days of treatment in healthy volunteers, and the assessment of GI and colonic transit by scintigraphy showed accelerated colonic transit and ascending colonic emptying [102].

Prucalopride (a dihydrobenzofuran-carboxamide derivative) is a selective and specific 5-HT₄ receptor agonist endowed with enterokinetic properties, recently approved by the European Medicines Agency (EMA) for treatment of idiopathic chronic constipation. Over a 12-week treatment period, 2 and 4 mg of prucalopride once daily significantly improved bowel habits in three large, randomized, double-blind, multicenter trials in patients with severe chronic constipation. Prucalopride was well tolerated: the incidence of QT interval prolongation was low and similar to that induced by placebo [97]. Prucalopride is able to penetrate the blood–brain barrier, and it binds to 5-HT₄ receptors in the rat brain [103].

Manini et al. [104] reported that the 5-HT₄ agonist velusetrag significantly accelerated intestinal and colonic transit (ascending colon emptying) after a single dose and accelerated gastric emptying after multiple dosing. Hoffman et al. [105] demonstrated that, similar to other 5-HT₄ receptor, velusetrag promotes propulsive motility and attenuates visceral hypersensitivity, but the precise mechanisms remain unclear.

Conclusions

In the CNS and in the gut, altered 5-HT signaling contributes to hypersensitivity and abnormal gut function in IBS, suggesting that this neurotransmitter with endocrine and paracrine functions may play a role in the development of IBS symptoms. The therapeutic effects of 5-HT₃ antagonists and 5-HT₄ agonists in IBS treatment are likely to be due also to the ability to modulate visceral nociception in the central stress circuit. It would be important to continue to investigate other receptor subtypes in the brain, particularly those regulating the affective states and the response to stress, in order to clarify their role in visceral perception. Further studies along these lines are needed in order to develop an optimal treatment for IBS.

Conflict of interest None.

References

1. Atkinson W, Lockhart S, Whorwell PJ, Keevil B, Houghton LA (2006) Altered 5-hydroxytryptamine signaling in patients with constipation-and diarrhea-predominant irritable bowel syndrome. *Gastroenterology* 130:34–43
2. Stasi C, Rosselli M, Bellini M et al (2012) Altered neuro-endocrine-immune pathways in the irritable bowel syndrome: the top-down and the bottom-up model. *J Gastroenterol* 47:1177–1185
3. Ford AC, Talley NJ, Schoenfeld PS et al (2009) Efficacy of antidepressants and psychological therapies in irritable bowel syndrome: systematic review and meta-analysis. *Gut* 58:367–378

4. Bearcroft CP, Perret D, Farthing MJ (1998) Postprandial plasma 5-hydroxytryptamine in diarrhea-predominant irritable bowel syndrome: a pilot study. *Gut* 42:42–46
5. Houghton LA, Foster JM, Whorwell PJ (2003) Alosetron, a 5-HT₃ receptor antagonist, delays colonic transit in patients with irritable bowel syndrome and healthy volunteers. *Aliment Pharmacol Ther* 14:775–782
6. Spiller RC, Jenkins D, Thornley JP (2000) Increased rectal mucosal entero-endocrine cells, T lymphocytes, and increased gut permeability following acute *Campylobacter* enteritis and in post-dysenteric irritable bowel syndrome. *Gut* 47:804–811
7. Dunlop SP, Coleman NS, Blackshaw E et al (2005) Abnormalities of 5-Hydroxytryptamine metabolism in irritable bowel syndrome. *Clin Gastroenterol Hepatol* 3:349–357
8. Kim DY, Camilleri M (2000) Serotonin: a mediator of the brain-gut connection. *Am J Gastroenterol* 95:2698–2709
9. Grundy D, Al-Chaer ED, Aziz Q et al (2006) Fundamentals of neurogastroenterology: basic science. *Gastroenterology* 130:1391–1411
10. Varnas K, Hallidin C, Hall H (2004) Autoradiographic distribution of serotonin transporters and receptor subtypes in human brain. *Hum Brain Mapp* 22:246–260
11. Kirchgessner AL, Liu MT, Raymond JR et al (1996) Identification of cells that express 5-hydroxytryptamine_{1A} receptors in the nervous systems of the bowel and pancreas. *J Comp Neurol* 364:439–455
12. Fiorica-Howells E, Hen R, Gingrich J et al (2002) 5-HT_{2A} receptors: location and functional analysis in intestines of wild-type and 5-HT_{2A} knockout mice. *Am J Physiol Gastrointest Liver Physiol* 282:G877–G893
13. Hoffman JM, Tyler K, MacEachern SJ et al (2012) Activation of colonic mucosal 5-HT₄ receptors accelerates propulsive motility and inhibits visceral hypersensitivity. *Gastroenterology* 142:844–854
14. Mawe GM, Branchek TA, Gershon MD (1986) Peripheral neural serotonin receptors: identification and characterization with specific antagonists and agonists. *Proc Natl Acad Sci USA* 83:9799–9803
15. Gershon MD (1999) Roles played by 5-hydroxytryptamine in the physiology of the bowel. *Aliment Pharmacol Ther* 13:15–30
16. Tonini M (2005) 5-Hydroxytryptamine effects in the gut: the 3, 4 and 7 receptors. *Neurogastroenterol Motil* 17:637–642
17. Pazos A, Prost TA, Palacios JM (1987) Serotonin receptors in the human brain III. Autoradiographic mapping of serotonin-1 receptors. *Neuroscience* 21:97–122
18. Pineyro G, Blier P (1999) Autoregulation of serotonin neurons: role in antidepressant drug action. *Pharmacol Rev* 51:533–591
19. Wang GD, Wang XY, Zou F et al (2013) Mast cell expression of the serotonin_{1A} receptor in guinea pig and human intestine. *Am J Physiol Gastrointest Liver Physiol* 304:G855–G863
20. Humprey PP, Hartig P, Hoyer D (1993) A proposed new nomenclature for 5-HT receptors. *Trends Pharmacol Sci* 14:233–236
21. Tepper SJ, Rapoport AM, Sheftell FD (2002) Mechanisms of action of the 5-HT_{1B/1D} receptor agonists. *Arch Neurol* 59:1084–1088
22. Hamel E, Fan E, Linville D et al (1993) Expression of mRNA for the serotonin 5-hydroxytryptamine_{1D} beta receptor subtype in human and bovine cerebral arteries. *Mol Pharmacol* 44:242–246
23. Levy FO, Gudermann T, Peres-Reyes E et al (1992) Molecular cloning of a human serotonin receptor (S12) with a pharmacological profile resembling that of the 5-HT_{1D} subtype. *J Biol Chem* 267:7553–7562
24. Jin H, Oksenberg D, Ashkenazi A et al (1992) Characterization of the human 5-hydroxytryptamine_{1B} receptor. *J Biol Chem* 267:5735–5738
25. Weinshank RL, Zgombick JM, Macchi MJ et al (1992) Human serotonin 1D receptor is encoded by a subfamily of two distinct genes: 5-HT_{1D α} and 5-HT_{1D β} . *Proc Natl Acad Sci USA* 89:3630–3634
26. Lowther S, De Paermentier F, Crompton MR et al (1992) The distribution of 5-HT_{1D} and 5-HT_{1E} binding sites in human brain. *Eur J Pharmacol* 222:137–142
27. McAllister G, Charlesworth A, Snodin C et al (1992) Molecular cloning of a serotonin receptor from human brain (5HT_{1E}): a fifth 5HT₁-like subtype. *Proc Natl Acad Sci USA* 89:5517–5521
28. Adham N, Kao HT, Schechter LE et al (1993) Cloning of another human serotonin receptor (5-HT_{1F}): a fifth 5-HT₁ receptor subtype coupled to the inhibition of adenylate cyclase. *Proc Natl Acad Sci USA* 90:408–412
29. Molderings GJ, Werner K, Likungu J et al (1990) Inhibition of noradrenaline release from the sympathetic nerves of the human saphenous vein via presynaptic 5-HT receptor similar to the 5-HT_{1D} subtype. *Naunyn Schmiedebergs Arch Pharmacol* 342:371–377
30. Bax WA, Renzenbrink GJ, Van Heuven-Nolsen D et al (1993) 5-HT receptors mediating contractions of the isolated human coronary artery. *Eur J Pharmacol* 239:203–210
31. Tack J, Coulter B, Wilmer A et al (1998) Actions of the 5-hydroxytryptamine 1 receptor agonist sumatriptan on interdigestive gastrointestinal motility in man. *Gut* 42:36–41
32. Foguet M, Hoyer D, Pardo LA et al (1992) Cloning and functional characterization of the rat stomach fundus serotonin receptor. *EMBO J* 11:3481–3487
33. Bubar MJ, Cunningham KA (2006) Serotonin 5-HT_{2A} and 5-HT_{2C} receptors as potential targets for modulation of psychostimulant use and dependence. *Curr Top Med Chem* 6:1971–1985
34. Vane JR (1959) The relative activities of some tryptamine analogues on the isolated rat stomach strip preparation. *Br J Pharmacol* 14:87–98
35. Cohen ML, Wittenauer LA (1987) Serotonin receptor activation of phosphoinositide turnover in uterine, fundal, vascular and tracheal smooth muscle. *J Cardiovasc Pharmacol* 10:176–181
36. Kursar JD, Nelson DL, Wainscott DB et al (1992) Molecular cloning, functional expression and pharmacological characterization of a novel serotonin receptor (5-hydroxytryptamine_{2F}) from rat stomach fundus. *Mol Pharmacol* 42:549–557
37. Kennet GA, Aisworth K, Trail B et al (1997) BW 723C86, a 5HT_{2B} receptor antagonist, causes hyperphagia and reduced grooming in rats. *Neuropharmacology* 36:23–39
38. Gershon MD (2003) Serotonin and its implication for the management of irritable bowel syndrome. *Rev Gastroenterol Disord* 3:S25–S34
39. Pasqualetti M, Ori M, Castagna M et al (1999) Distribution and cellular localization of the serotonin type 2C receptor messenger RNA in human brain. *Neuroscience* 92:601–611
40. Boess FG, Martin IL (1994) Molecular biology of 5HT receptors. *Neuropharmacology* 33:275–317
41. Pratt GD, Bowery NG, Kilpatrick GJ et al (1990) Consensus meeting agrees distribution of 5-HT₃ receptors in mammalian hindbrain. *Trends Pharmacol Sci* 11:135–137
42. Thompson AJ, Lummis SC (2007) The 5-HT₃ receptor as a therapeutic target. *Expert Opin Ther Targets* 11:527–540
43. Kidd EJ, Laporte AM, Langlois X et al (1993) 5-HT₃ receptors in the rat central nervous system are mainly located on nerve fibres and terminals. *Brain Res* 612:289–298
44. Hamon M, Gallissot MC, Menard F et al (1989) 5-HT₃ receptor binding sites are on capsaicin-sensitive fibres in the rat spinal cord. *Eur J Pharmacol* 164:315–322
45. Takaki M (2003) Gut pacemaker cells: the interstitial cells of Cajal. *J Smooth Muscle Res* 39:137–161

46. Liu HN, Ohya S, Nishizawa Y et al (2011) Serotonin augments gut pacemaker activity via 5-HT₃ receptors. *PLoS ONE* 6:e24928
47. Marchetti E, Dumuis A, Bockaert J et al (2000) Differential modulation of the 5-HT₄ receptor agonists and antagonist on rat learning and memory. *Neuropharmacology* 39:2017–2027
48. King MV, Marsden CA, Fone KC (2008) A role for the 5-HT_{1A}, 5-HT₄ and 5-HT₆ receptors in learning and memory. *Trends Pharmacol Sci* 29:482–492
49. Salmon E (2007) A review of the literature on neuroimaging of serotonergic function in Alzheimer's disease and related disorders. *J Neural Transm* 114:1179–1185
50. Bockaert J, Claeysen S, Compan V et al (2011) 5-HT₄ receptors, a place in the sun: act two. *Curr Opin Pharmacol* 11:87–93
51. Madsen K, Haahr MT, Marnier L et al (2011) Age and sex effects on 5-HT₄ receptors in the human brain: a [(11C)SB207145 PET study. *J Cereb Blood Flow Metab* 31:1475–1481
52. Tack J, Camilleri M, Chang L et al (2012) Systematic review: cardiovascular safety profile of 5-HT₄ agonists developed for gastrointestinal disorders. *Aliment Pharmacol Ther* 35:745–767
53. Schoemaker RG, Du XY, Bax WA et al (1993) 5-Hydroxytryptamine stimulates human isolated atrium but not ventricle. *Eur J Pharmacol* 239:103–105
54. Oliver KR, Kinsey AM, Wainwright A et al (2000) Localization of 5-HT_{5A} receptor-like immunoreactivity in the rat brain. *Brain Res* 867:131–142
55. Pasqualetti M, Ori M, Nardi I et al (1998) Distribution of the 5-HT_{5A} serotonin receptor mRNA in the human brain. *Mol Brain Res* 56:1–8
56. Plassat J-L, Boschert U, Amlaiky N et al (1992) The mouse 5-HT_{5A} receptor reveals a remarkable heterogeneity within the 5-HT_{1D} receptor family. *EMBO J* 11:4779–4786
57. Grailhe R, Grabtree GW, Hen R (2001) Human 5-HT₅ receptors: the 5-HT_{5A} receptor is functional but the 5-HT_{5B} receptor was lost during mammalian evolution. *Eur J Pharmacol* 418:157–167
58. Wesolowski A (2002) In the search for selective ligands of 5-HT₅, 5-HT₆ and 5-HT₇ serotonin receptors. *Pol J Pharmacol* 54:327–341
59. Thomas DR, Hagan JJ (2004) 5-HT₇ receptors. *Curr Drug Targets CNS Neurol Disord* 3:81–90
60. Tonini M, Vicini R, Cervio E et al (2005) 5-HT₇ receptors modulate peristalsis and accommodation in the guinea pig ileum. *Gastroenterology* 129:1557–1566
61. Bühlbring E, Lin RC (1985) The effect of intraluminal application of 5-hydroxytryptamine and 5-hydroxytryptophan on peristalsis; the local production of 5-HT and its release in relation to intraluminal pressure and propulsive activity. *J Physiol* 140:381–407
62. Grider JR (1994) CGRP as a transmitter in the sensory pathway mediating peristaltic reflex. *Am J Physiol* 266:G1139–G1145
63. Danzebrink RM, Gebhart GF (1991) Evidence that spinal 5-HT₁, 5-HT₂ and 5-HT₃ receptor subtypes modulate responses to noxious colorectal distension in the rat. *Brain Res* 538:64–75
64. Miftahof R, Akhmadeev NR (2007) Neurochemical bases of visceral nociception: mathematical model. *J Theor Biol* 249:343–360
65. Almeida TF, Roizenblatt S, Tufik S (2004) Afferent pain pathways: a neuroanatomical review. *Brain Res* 1000:40–56
66. Camilleri M, Coulie B, Tack JF (2001) Visceral hypersensitivity: facts, speculations, and challenges. *Gut* 48:125–131
67. Bueno L, Fioramonti J, Delvaux M et al (1997) Mediators and pharmacology of visceral sensitivity: from basic to clinical investigations. *Gastroenterology* 112:1714–1743
68. Keszhelyi D, Troost FJ, Jonkers DM et al (2013) Decreased levels of kynurenic acid in the intestinal mucosa of IBS patients: relation to serotonin and psychological state. *J Psychosom Res* 74:501–504
69. Stasi C, Bellini M, Costa F et al (2013) Neuroendocrine markers and psychological features in patients with irritable bowel syndrome. *Int J Colorectal Dis* 28:1203–1208
70. Muller-Lissner SA, Bollani S, Brummer RJ et al (2001) Epidemiological aspects of irritable bowel syndrome in Europe and North America. *Digestion* 64:200–204
71. Nakai A, Kumakura Y, Boivin M et al (2003) Sex differences of brain serotonin synthesis in patients with irritable bowel syndrome using alpha-[¹¹C]methyl-L-tryptophan, positron emission tomography and statistical parametric mapping. *Can J Gastroenterol* 17:191–196
72. Creed F, Fernandes L, Guthrie E et al (2003) The cost-effectiveness of psychotherapy and paroxetine for severe irritable bowel syndrome. *Gastroenterology* 124:303–317
73. Creed F, Tomenson B, Guthrie E et al (2008) The relationship between somatisation and outcome in patients with severe irritable bowel syndrome. *J Psychosom Res* 64:613–620
74. Mayer EA, Naliboff BD, Chang L (2001) Basic pathophysiologic mechanisms in irritable bowel syndrome. *Dig Dis* 19:212–218
75. Mertz H (2002) Role of the brain and sensory pathways in gastrointestinal sensory disorders in humans. *Gastroenterology* 122:i29–i33
76. Whitehead WE, Engel BT, Schuster MM (1980) Irritable bowel syndrome: physiological and psychological differences between diarrhea-predominant and constipation-predominant patients. *Dig Dis Sci* 25:404–413
77. Prior A, Maxton DG, Whorwell PJ (1990) Anorectal manometry in irritable bowel syndrome: differences between diarrhoea and constipation predominant subjects. *Gut* 31:458–462
78. Simrén M, Abrahamsson H, Björnsson ES (2001) An exaggerated sensory component of the gastrocolonic response in patients with irritable bowel syndrome. *Gut* 48:20–27
79. Slater BJ, Plusa SM, Smith AN, Varma JS (1997) Rectal hypersensitivity in the irritable bowel syndrome. *Int J Colorectal Dis* 12:29–32
80. Harraf F, Schmulson M, Saba L et al (1998) Subtypes of constipation predominant irritable bowel syndrome based on rectal perception. *Gut* 43:388–394
81. Steens J, Van Der Schaar PJ, Penning C et al (2002) Compliance, tone and sensitivity of the rectum in different subtypes of irritable bowel syndrome. *Neurogastroenterol Motil* 14:241–247
82. Drossman DA, Ringel Y, Vogt BA et al (2003) Alterations of brain activity associated with resolution of emotional distress and pain in a case of severe irritable bowel syndrome. *Gastroenterology* 124:754–761
83. Baciú MV, Bonaz BL, Papillon E et al (1999) Central processing of rectal pain: a functional MR imaging study. *Am J Neuroradiol* 20:1920–1924
84. Bonaz B, Baciú M, Papillon E et al (2002) Central processing of rectal pain in patients with irritable bowel syndrome: an fMRI study. *Am J Gastroenterol* 97:654–661
85. Suzuki R, Rygh LJ, Dickenson AH (2004) Bad news from the brain: descending 5-HT pathways that control spinal pain processing. *Trends Pharmacol Sci* 25:613–617
86. Sharma HS, Cervos-Navarro J, Dey PK (1991) Increased blood-brain barrier permeability following acute short-term swimming exercise in conscious normotensive young rats. *Neurosci Res* 10:211–221
87. Oatway MA, Chen Y, Weaver LC (2004) The 5-HT₃ receptor facilitates at-level mechanical allodynia following spinal cord injury. *Pain* 110:259–268
88. Borman RA, Tilford NS, Harmen DW et al (2002) 5-HT_{2B} receptors play a key role in mediating the excitatory effects of 5-HT in human colon in vitro. *Br J Pharmacol* 135:1141–1151
89. Beattie DT, Smith JA, Marquess D et al (2004) The 5-HT₄ receptor agonist, tegaserod, is a potent 5-HT_{2B} receptor antagonist in vitro and in vivo. *Br J Pharmacol* 143:549–560

90. Gore S, Gilmore IT, Haigh CG et al (1990) Colonic transit in man is slowed by ondansetron (GR38032F), a selective 5-hydroxytryptamine receptor (type 3) antagonist. *Aliment Pharmacol Ther* 4:139–144
91. Steadman CJ, Talley NJ, Phillips SF et al (1992) Selective 5-hydroxytryptamine type 3 receptor antagonism with ondansetron as treatment for diarrhea-predominant irritable bowel syndrome: a pilot study. *Mayo Clin Proc* 67:732–738
92. Goldberg PA, Kamm MA, Setti-Carraro P et al (1996) Modification of visceral sensitivity and pain in irritable bowel syndrome by 5-HT₃ antagonism (ondansetron). *Digestion* 57:478–483
93. Viramontes BE, Camilleri M, McKinzie S et al (2001) Gender-related differences in slowing colonic transit by a 5-HT₃ antagonist in subjects with diarrhea-predominant irritable bowel syndrome. *Am J Gastroenterol* 96:2671–2676
94. Bharucha AE, Camilleri M, Haydock S et al (2000) Effects of a serotonin 5-HT₄ receptor antagonist SB-207266 on gastrointestinal motor and sensory function in humans. *Gut* 47:667–674
95. De Ponti F, Tonini M (2001) Irritable bowel syndrome: new agents targeting serotonin receptor subtypes. *Drugs* 61:317–332
96. Camilleri M (2001) Review article: tegaserod. *Aliment Pharmacol Ther* 15:277–289
97. Frampton JE (2009) Prucalopride. *Drugs* 69:2463–2476
98. Kozłowski CM, Green A, Grundy D et al (2000) The 5-HT₃ receptor antagonist alosetron inhibits the colorectal distention induced depressor response and spinal c-fos expression in the anaesthetised rat. *Gut* 46:474–480
99. Lee KJ, Kim NY, Kwon JK et al (2011) Efficacy of ramosetron in the treatment of male patients with irritable bowel syndrome with diarrhea: a multicenter, randomized clinical trial, compared with mebeverine. *Neurogastroenterol Motil* 23:1098–1104
100. Schikowski A, Thewissen M, Mathis C et al (2002) Serotonin type-4 receptors modulate the sensitivity of intramural mechanoreceptive afferents of the cat rectum. *Neurogastroenterol Motil* 14:221–227
101. Evans BW, Clark WK, Moore DJ et al (2007) Tegaserod for the treatment of irritable bowel syndrome and chronic constipation. *Cochrane Database Syst Rev* 4:003960
102. Camilleri M, Vazquez-Roque MI, Burton D et al (2007) Pharmacodynamic effects of a novel prokinetic 5-HT receptor agonist, ATI-7505, in humans. *Neurogastroenterol Motil* 19:30–38
103. Johnson DE, Drummond E, Grimwood S et al (2012) The 5-hydroxytryptamine₄ receptor agonists prucalopride and PRX-03140 increase acetylcholine and histamine levels in the rat prefrontal cortex and the power of stimulated hippocampal θ oscillations. *J Pharmacol Exp Ther* 341:681–691
104. Manini ML, Camilleri M, Goldberg M et al (2010) Effects of Velusetrag (TD-5108) on gastrointestinal transit and bowel function in health and pharmacokinetics in health and constipation. *Neurogastroenterol Motil* 22(42–49):e7–e8
105. Hoffman JM, Tyler K, MacEachern SJ et al (2012) Activation of colonic mucosal 5-HT₄ receptors accelerates propulsive motility and inhibits visceral hypersensitivity. *Gastroenterology* 142(844–854):e4